P-46

Development of 99Mtc-Labeling Protocol for Hydrogel-Based Microspheres

NIKOLETT HEGEDŰS¹; KRISZTIÁN SZIGETI¹; DOMOKOS MÁTHɹ²

¹ Department of Biophysics and Radiation Biology, Semmelweis University, Tűzoltó utca 37-47., H-1094 Budapest, Hungary ² CROmed Translational Research Centers, Baross utca 91-95., H-1047 Budapest, Hungary

Correspondence: hegedus.nikolett@med.semmelweis-univ.hu

Keywords: Microspheres, polyethylene glycol, ^{99m}Tc labeling, trans-arterial embolization agent, thin layer chromatography

1. INTRODUCTION

Hydrogel microspheres are biocompatible, hydrophilic and non-resorbable polymeric systems which are increasingly used as biomedical carriers to target and deliver diagnostic and therapeutic agents.¹ These microspheres usually consist of a swollen network of hydrophilic polymer chain with water content exceeding 95%, which guarantees elasticity, compressibility and stability. The conjugated groups are responsible for the carrier feature of the particles.²

The aim of this study was to develop an easy and effective protocol for radiolabeling LifePearl® microspheres with ^{99m}Tc-pertechnetate. LifePearl® is produced from polyethylene glycol chain and additional sulfonate groups which enable the loading of the beads with chemotherapeutic agents.³ This drug delivery system has been applied in FDA-approved manner as a trans-arterial chemoembolization (TACE) agent for the treatment of unresectable hepatocellular carcinoma.

2. MATERIALS AND METHODS

For radiolabeling procedure LifePearl® microspheres (Terumo European Interventional Systems, Leuven, Belgium; size range 100 ± 25 um) were used. The labeling process started with the chemical reduction of ^{99m}Tc-pertechnetate (^{99m}TcO₄; *Synthesis route No1*). For this purpose, freshly made special 'Reducing agent' was composed from 0.1% stannous-chloride, 0.2% ascorbic acid and concentrated hydrochloric acid for pH adjustment (pH 4-5). Next, the microspheres were incubated with the radioactive solution (30-50 MBq; mixing ratio varied between 1:5 to 5:1) at room temperature for various incubation time (10 min – 1 h). Following the auto-sedimentation of the gellike ^{99m}Tc-microspheres the supernatant was dis-

carded and the labeling efficiency, the retention factors (R_f) of the particles were determined by using instant thin layer chromatography (ITLC). For quality control (QC), instant thin-layer chromatography paper impregnated with silica gel (ITLC-SG; Agilent Technologies, Santa Clara, US) was used as stationary phase and 0.9% Saline was applied as solvent (mobile phase). The reading out of chromatographic paper was made by miniGITA scanning device (Elysia-raytest, Straubenhardt,

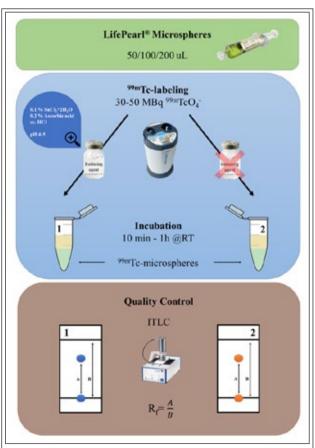


Figure 1 The synthesis scheme of ^{99m}Tc-labeled microspheres. (1) Labeling with 'Reducing-agent'. (2) Labeling without 'Reducing-agent'.

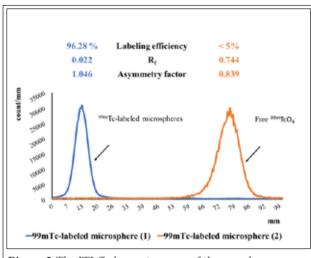


Figure 2 The ITLC chromatograms of the sample

Germany). The whole labeling procedure and quality control process were repeated without the application of 'Reducing agent' as negative control (*Synthesis route No2*) (see *Figure 1*).

3. RESULTS

The radiolabeling efficiency exceeds 95 % by *Synthesis route No1* (blue curve) and remained constant for several hours. It was independent from the applied microsphere concentration. In contrast, labeling with the unreduced form of ^{99m}Tc-pertechnetate (*Synthesis route No2; orange curve*), as

a negative control, resulted negligible labeling efficiency (less than 5%) (*Figure 2*).

4. CONCLUSION

The developed labeling protocol of LifePearl® microspheres is an easily applicable method to monitor the biodistribution of particles applied in oncologic therapy and to determine critical biodistribution of TACE agents.

5. ACKNOWLEDGEMENTS

The project was supported by Higher Education Institutional Excellence Program of Ministry of Human Capacities Hungary, within therapy theme of Semmelweis University.

References

- 1. de Baere T, Plotkin S, Yu R, Sutter A, Wu Y, Cruise GM. An in vitro evaluation of four types of drug-eluting microspheres loaded with doxorubicin, J. of Vasc. and Interv. Radiol., 27:1425-1431 (2016).
- 2. Laurent A, Velzenberger E, Wassef M, Pelage JP, Lewis AL. Do microspheres with narrow or standard size distributions localize differently in vasculature? An experimental study in sheep kidney and uterus, J. Vasc. Interv. Radiol., 19: 1733–1739 (2008).
- 3. Boulin M, Guiu S, Chauffert B, Aho S, Cercueil JP, Ghiring-helli F, Krause D, Fagnoni P, Hillon P, Bedenne L, Guiu B. Screening of anticancer drugs for chemoembolization of hepatocellular carcinoma, Anti-cancer drugs., 22: 741-8 (2011).